

Product Information

Cosmid Construction Kit-2

Research use only

Catalog No: R-1002

Lot No: 1016

Contents:

- Maltose/MgSO₄ mix, 0.7 mL (# R-1023)
- SM medium, 1 mL (# R-1004)
- Lambda packaging extract, 50 µL (#R-1005)
- OD101 glycerol stock, 200 µL (#R-1006)
- Amp/Kan antibiotic mix, 500 µL (#R-1007)
- Sfil enzyme, 70 µL (#R-1008)
- 10x Sfil buffer, 200 µL (#R-1009)

Storage: -20°C for the Sfil enzyme and 10x Sfil buffer; -70 °C for all other reagents.

Features and Applications: These reagents are used to construct at least 10 AdenoQuick2.0 cosmids.

- ◆ Sfil is used to digest the 4 adenovirus shuttle plasmids.
- ◆ Maltose and MgSO₄ are added to the *E. coli* OD101 culture to induce the expression of the receptor for phage λ.
- ◆ The lambda packaging extract is used to package the cosmid ligation products into phage λ.
- ◆ The SM medium is used to dilute the packaging reaction at the end of the 30-minutes incubation period.
- ◆ OD101 is the *E coli* strain that will be infected with the packaged phage λ.

Note: All reagents are sterile and must be handled accordingly.
Quick protocol

Preparation of λ-competent *E. coli*

- Add 2 mL LB + 60 µL MgSO₄/maltose solution in a sterile 10-mL tube.
- Inoculate with one colony of an overnight culture of OD101.
- Grow at 37 °C until the culture appears cloudy.
- Chill on ice.
- Dilute the cells with LB to Abs_{600nm} = 1

Sfil Digestion

- Digest your 4 shuttle plasmids (0.5 µg each) with Sfil in a single vial for 30 min at 50°C.

Ligation

- Mix 8 µL Sfil-digested plasmid mix + 1 µL T4 ligase + 1 µl 10x T4 ligase buffer.
- Incubate for 1 hr at room temperature for 1 hr.

Packaging

- To a microcentrifuge tube, add:
 - 2 µL ligation mix

- 5 µL Lambda packaging extract
- Incubate at 30°C for 30 minutes.
- Add 100 µL SM medium.
- Add 5 µL chloroform.
- Mix the tube until a white precipitate appears.
- Spin at 13,000 rpm for 20 sec.
- Transfer the supernatant containing the packaged phage λ to a clean tube.

Infection and clone selection

- Mix 60 µL packaged phage λ and 60 µL diluted competent cells.
- Incubate for 30 min at 37 °C.
- Add 20 µL Amp/Kan antibiotic mix and spread on a Petri dish containing ~ 20 mL LB/agar.
- Incubate overnight at 37°C.

Consult the AdenoQuick manual for details.

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